

Update on the Tilapia Rickettsia Like Organism (TRLLO) Infecting Tilapia On Oahu.

Clyde S. Tamaru, Ph.D., Jim Brock, DVM, Nathene Lynn Antonio, Allen Riggs, DVM, Kathleen McGovern-Hopkins, Ruth Ellen Klinger-Bowen, Bradley Fox, Ph.D.

CTAHR/HAAA Workshop
Windward Community College
November 12, 2011



REGIONAL BIOSECURITY: OPERATIONAL BIOSECURITY AND DIAGNOSTIC SURVEILLANCE – Year 2

- Grant #: 2008-202(b)
- Reporting Period: April 30, 2011 – May 1, 2012.
- Funding Level: \$50,000
- Participants:
 - Shemiah Iaea
 - Ilima-Ho Lastimoso
 - Leina’ala Bright
 - Stan Kodama
 - Ron Koza



College of Tropical
Agriculture and Human
Resources



Aquaculture & Livestock
Support Services
DEPARTMENT OF AGRICULTURE



College of Tropical Agriculture and Human Resources
University of Hawai'i at Mānoa

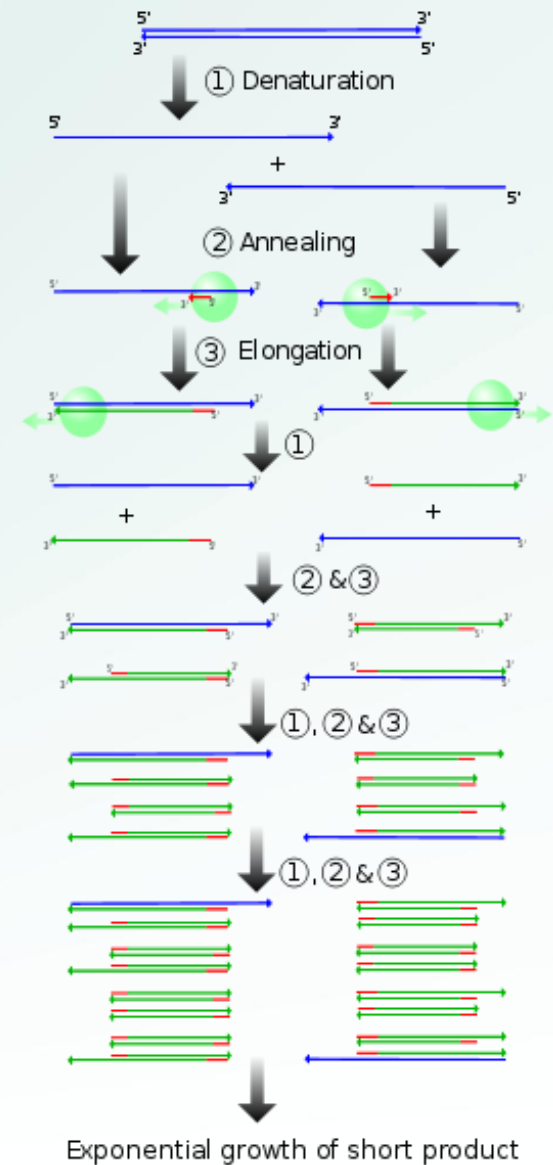
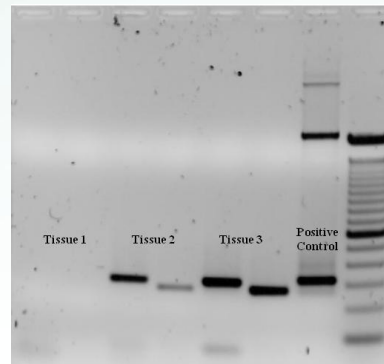
Rationale for Project

- In 1994, wild and farm-raised populations of tilapia on Oahu began to die of an unknown disease.
- Only tilapia were affected and mortalities occurred only during the cooler months (October to April) and were restricted to Oahu.
- Department of Agriculture issues PQ Policy 98-09, Section 150A-8, HRS effective November 5, 1998: Oahu shipments presented to PQB should not be certified for movement to other islands.
- Development and widespread availability of molecular detection techniques provides an opportunity to address the challenges posed by this particular pathogen.



Polymerase Chain Reaction (PCR)

The polymerase chain reaction (PCR) is a technique in molecular biology that amplifies a piece of DNA by generating millions of copies of a particular DNA sequence.



Primer's for TRLO/FLB

FLB16S180f: 5'-GCG-GAT-TAA- AGG-TGG-CCT-TTG-3' (forward primer)

FLB16S465r: 5'-CCT-GCA-AGC-TAT-TAA-CTC-ACA- 3' (reverse primer).

Hsieh et.al., 2007. PCR and in situ hybridization for the detection and localization of a new pathogen *Francisella*-like bacterium (FLB) in ornamental cichlids. *Diseases of Aquatic Organisms* **75**, 29-36.

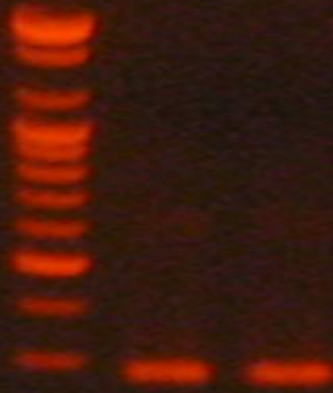
Misumi, et al., 2010 Identification and isolation of *Francisella*-like bacteria (FLB) from tilapia (*Oreochromis mossambicus*) for the first time in Hawaii. Submitted to Fisheries Science.



Validation of PCR Test for TRLO/FLB

Primers: TRLO F/R

Lane 1: Ladder
Lane 2: 10-87 Tilapia spleen
Lane 3: 10-87 Tilapia spleen
Lane 4: no template



TRLO positive sample from
Dr Riggs (ADP case # 10-87)

Primers: CO-I (Fish DNA)

Lane 1: Ladder
Lane 2: 10-87 Tilapia spleen
Lane 3: 10-87 Tilapia spleen
Lane 4: no template



PCR Validation: DNA Sequence

10-87 (+TRLO)

GGATCTACTGCGTTGGATAGCTAGTTGGTGGGGTAAGGGCCTACCAAGGCTACGATCCATA
GCTGATTTGAGAGGATGATCAGCCACATTGGGACTGAGACACGGCCCAAACCTCCTACGGG
AGGCAGCAGTGGGGAATATTGGACAATGGGGGAAACCCTGATCCAGCAATGCCATGTGTG
TGAAGAAGGCTCTAGGGTTGTAAAGCACTTTAGTTGGGGAGGAAAGCCTGTGAGTTATAG
CTTGCAGGAA

MTI-10-22C1 (+CO-I fish DNA)

ACGATCTGCATTGGTGCTTGAGCCGGATAGTAGGACTGCACTAAGTCTCCTTATTCGGGCA
GAACTAAGCCAGCCCGGCTCTCTCCTCGGAGACGACCAGATTTATAATGTAATTGTTACAG
CACATGCTTTCGTAATAATTTTCTTTATAGTAATACCAATTATAATTGGAGGGTTTGGAAACT
GACTGGTGCCACTTATGATTGGAGCACCAGACATGGCCTTCCCTCGAATAAATAACATGAG
TTTTTGACTCCTTCCCCCCTCATTCTCCTTCTCCTCGCCTCATCCGGAGTCGAAGCAGGA
GCCGGTACAGGATGAACTGTTTATCCTCCCCTCGCAGGCAATCTCGCCCACGCTGGACCTT
CTGTTGACTTGACCATCTTTTCCCTCCACTTGGCCGGGGTGTCATCTATTTTAGGCGCAATT
AATTTTATCACAACCATTATTAACATAAAACCCCCTGCCATCTCTCAATATCAAACACCCCT
CTTTGTGTGATCCGTTCTAATTACCGCAGTATTACTCCTACTATCCCTGCCCGTTCTTGCCGC
CGGCATCACAATACATCTAACAGATCGAAACCTGGACGCAAGCTTCTCTCACCCCTGCCAG
AAAAAGAGACTC



PCR Validation: TRLO = *Francisella* like bacteria (FLB) DNA Sequence

Descriptions

Send for links to other resources: [U](#) UniGene [E](#) OEO [G](#) Gene [S](#) Structure [M](#) Map Viewer [P](#) PubChem BioAssay

Sequences producing significant alignments:

Accession	Description	Max score	Total score	Query coverage	E value	Max ident	Links
FJ217162.1	<i>Francisella noatunensis</i> subsp. <i>orientalis</i> strain Ind04 16S ribosomal RNA g	552	442	97%	5e-121	99%	
EU682020.1	<i>Francisella noatunensis</i> subsp. <i>orientalis</i> strain Ehime-1 16S ribosomal RNA	552	442	97%	5e-121	99%	
EU672694.1	<i>Francisella</i> sp. LADL 07-285A 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
EU592612.1	<i>Francisella</i> sp. PQ1104 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AY282222.1	<i>Francisella</i> sp. AF-03-27 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AY282222.1	<i>Francisella</i> sp. AF-01-28 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AY282221.1	<i>Francisella</i> sp. AF-01-27 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AY282222.1	<i>Francisella</i> sp. AF-01-6 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AY282228.1	<i>Francisella</i> sp. AF-01-2 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
GQ2021425.1	<i>Francisella</i> sp. AF-04-405 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
GQ2021424.1	<i>Francisella</i> sp. AF-04-15 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
GQ2021423.1	<i>Francisella</i> sp. AF-03-28 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AF206675.1	Tilapia parasite TPT-541 16S ribosomal RNA gene, partial sequence	552	442	97%	5e-121	99%	
AF282827.2	<i>Cf. Francisella</i> sp. CYH-2002 16S ribosomal RNA gene, partial sequence; 16	552	442	97%	5e-121	99%	
GQ2171277.1	Uncultured <i>Francisella</i> sp. done KST1 16S ribosomal RNA gene, partial seq.	552	442	97%	5e-121	99%	
AB124068.1	<i>Francisella</i> sp. Ehime-1 gene for 16S rRNA, partial sequence	552	442	97%	5e-121	99%	

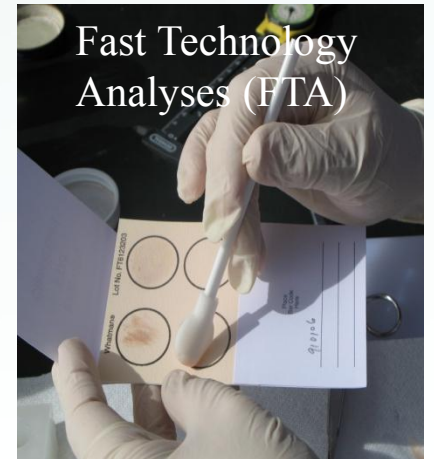
rt

NCBI Blast Nucleotide... Sequencing Results Bl... Positive Control (TRL...

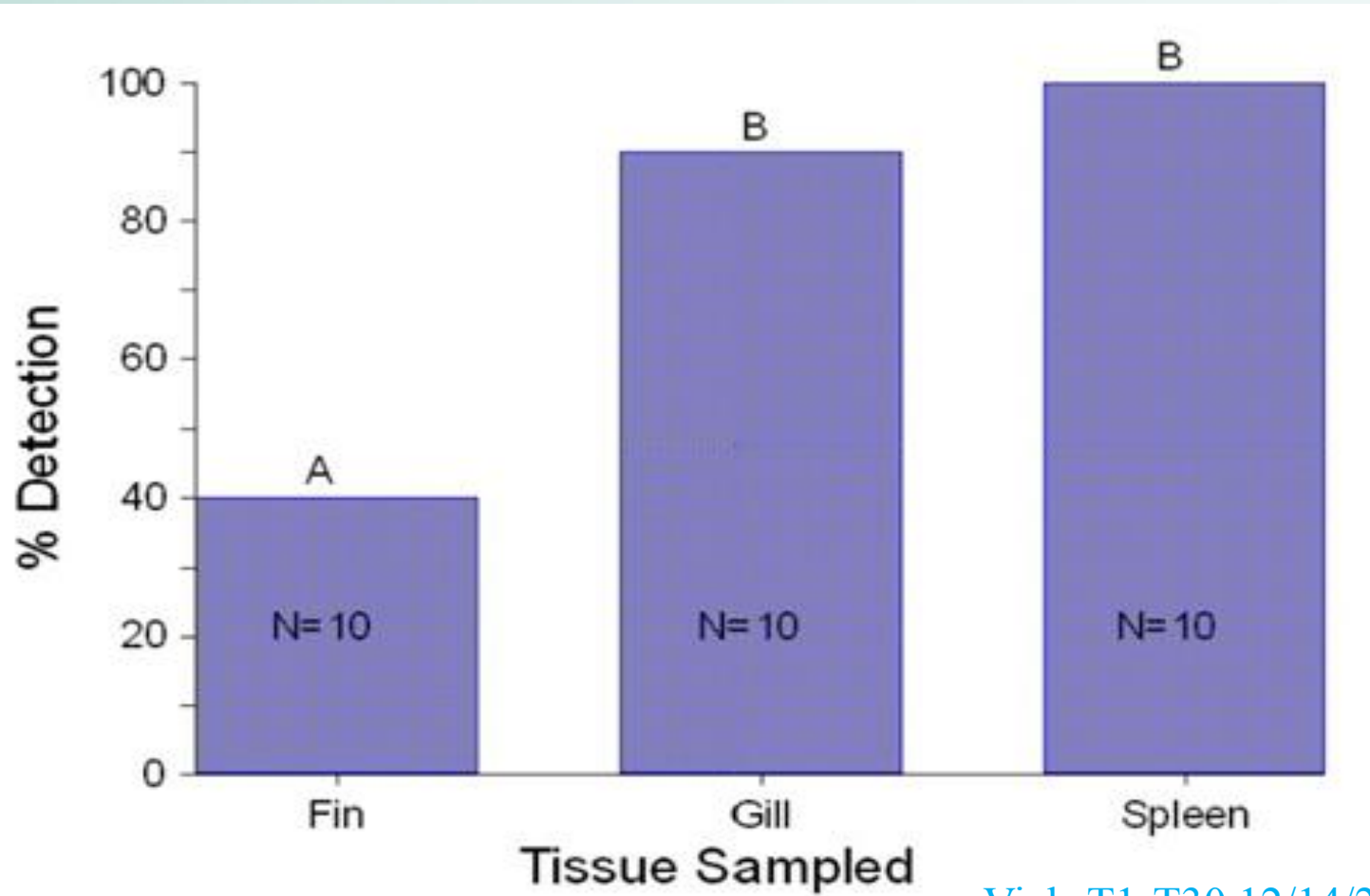
Basic Local Alignment Search Tool (BLAST)



Processing a specimen for PCR testing



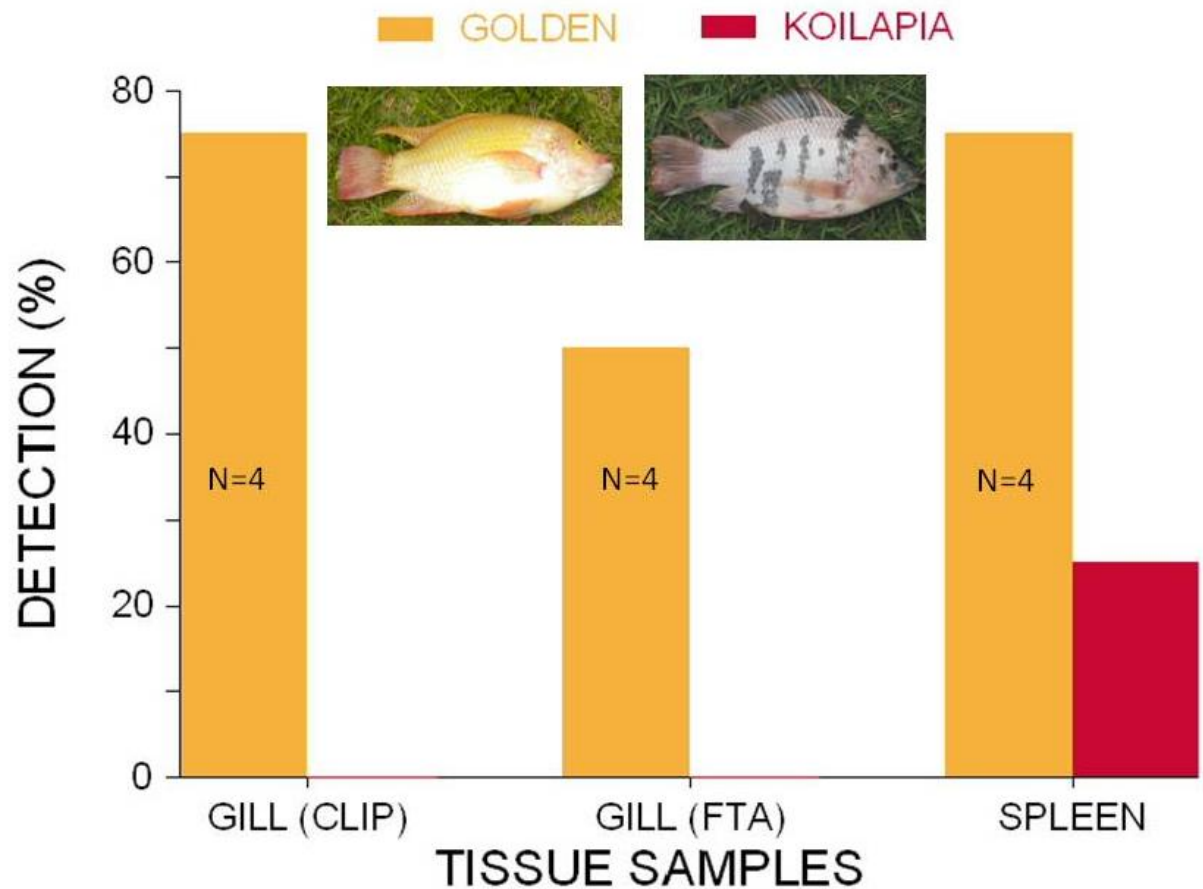
Tissue distribution during an active infection



Vials T1-T30 12/14/2010



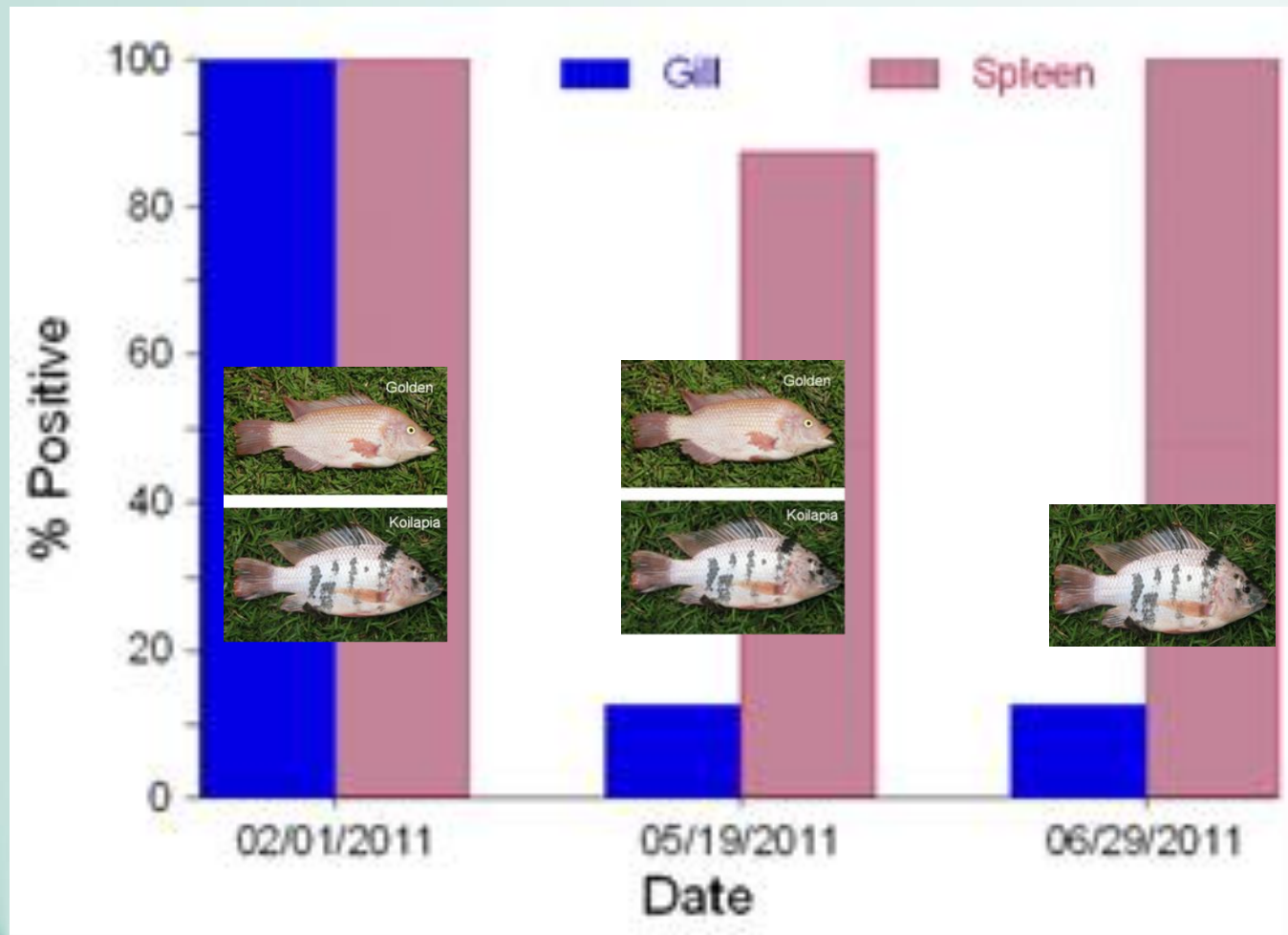
Species and tissue distribution of TRLO/FLB during an active infection



Vials T79 – T120 03/02/2011

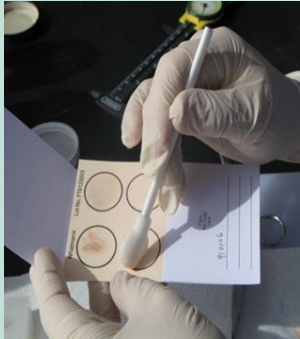


Change in tissue distribution of TRLO/FLB during active infection that becomes an asymptomatic population



Tissue distribution of TRLO/FLB in an asymptomatic population

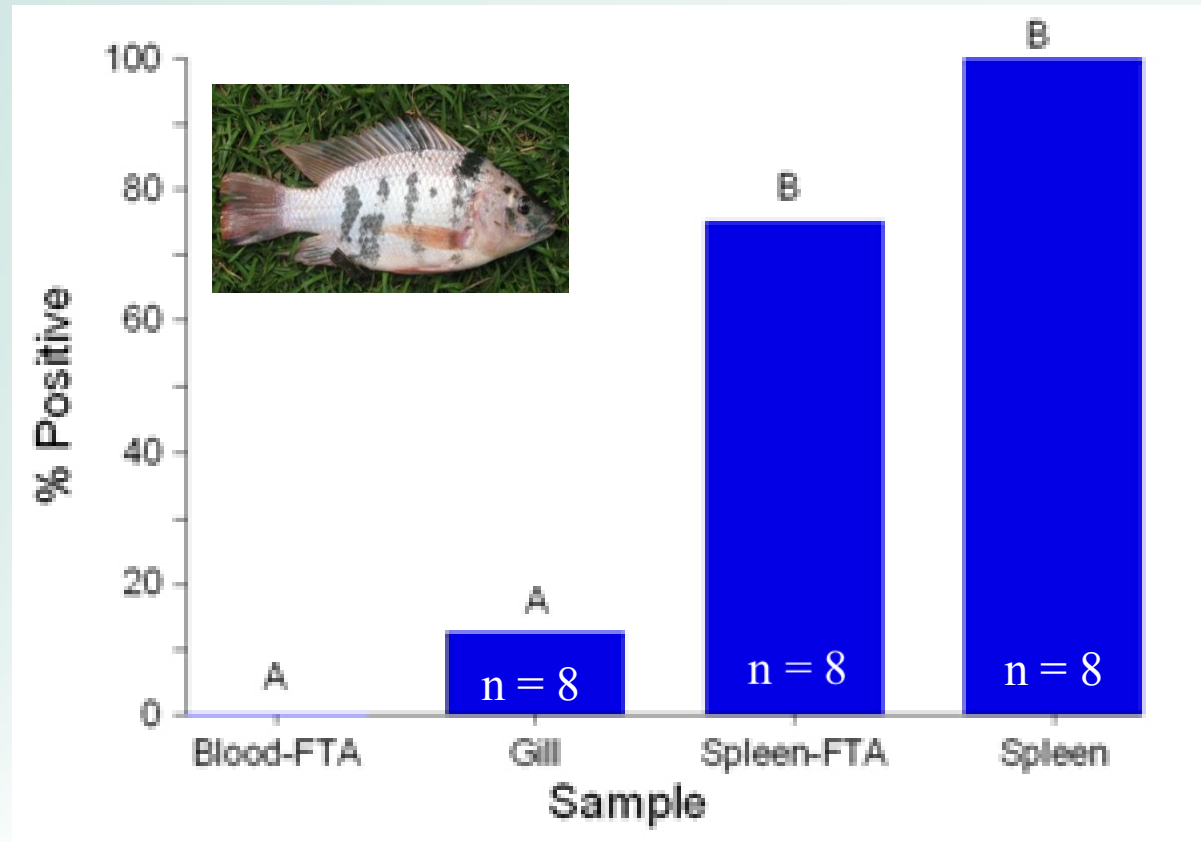
FTA
Card



Gill
Non-
lethal



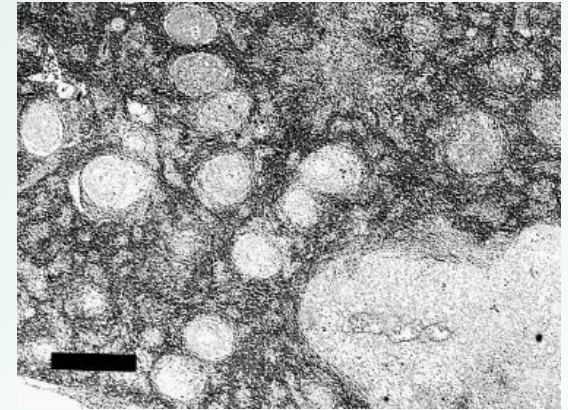
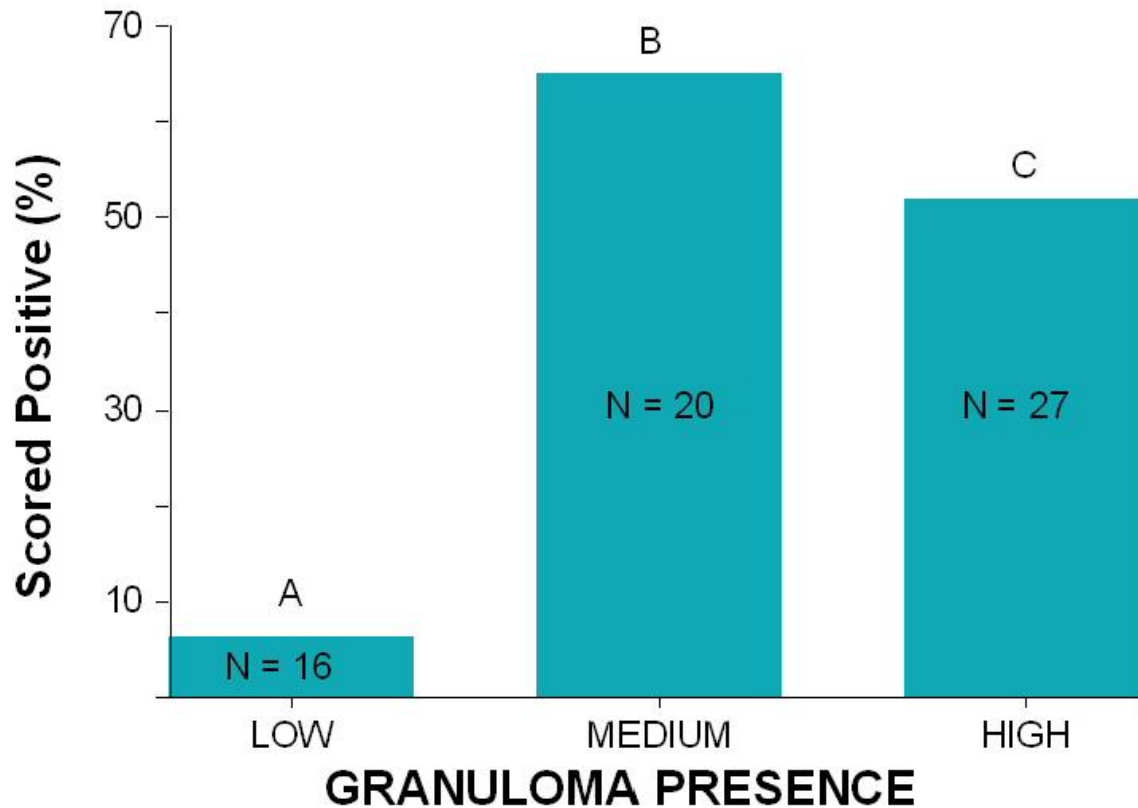
Spleen-
lethal



Vials T-143 - T174 06/29/2011



Use of granulomas as an indicator of TRLO/FLB




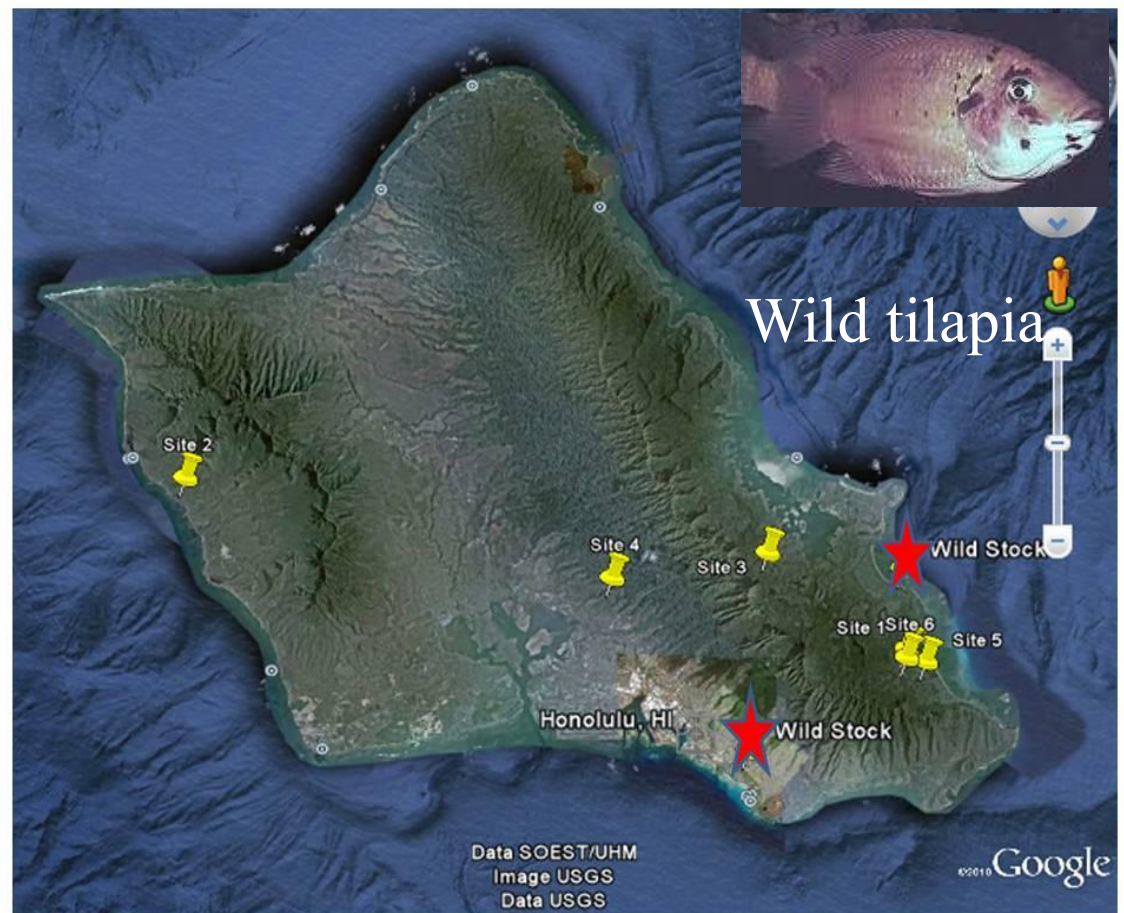
H&E stained spleen showing multiple granulomas. Scale bar = 50 μ m.

From Mauel, et al., 2005. *J. Vet. Diag. Invest.* 17:601-605.



Distribution of positive cases on Oahu Dec 2010 – Current

- Six cases from cultured stock 
- Two cases from wild stocks 
- More being tested



Summary and Conclusions

- Capacity for PCR testing of FLB (previously known as TRLO) established, validated and operational
- Several tissues can harbor FLB during an active infection
- Only spleen and lethal sampling protocol useful for identifying asymptomatic carriers
- Some tilapia strains may be resistant to FLB



MAHALO and Thanks for Listening

